



Anaplasma and *Ehrlichia* infections in wildlife reservoirs and livestock in Shimba Hills National Reserve (SHNR), Kwale County, Kenya

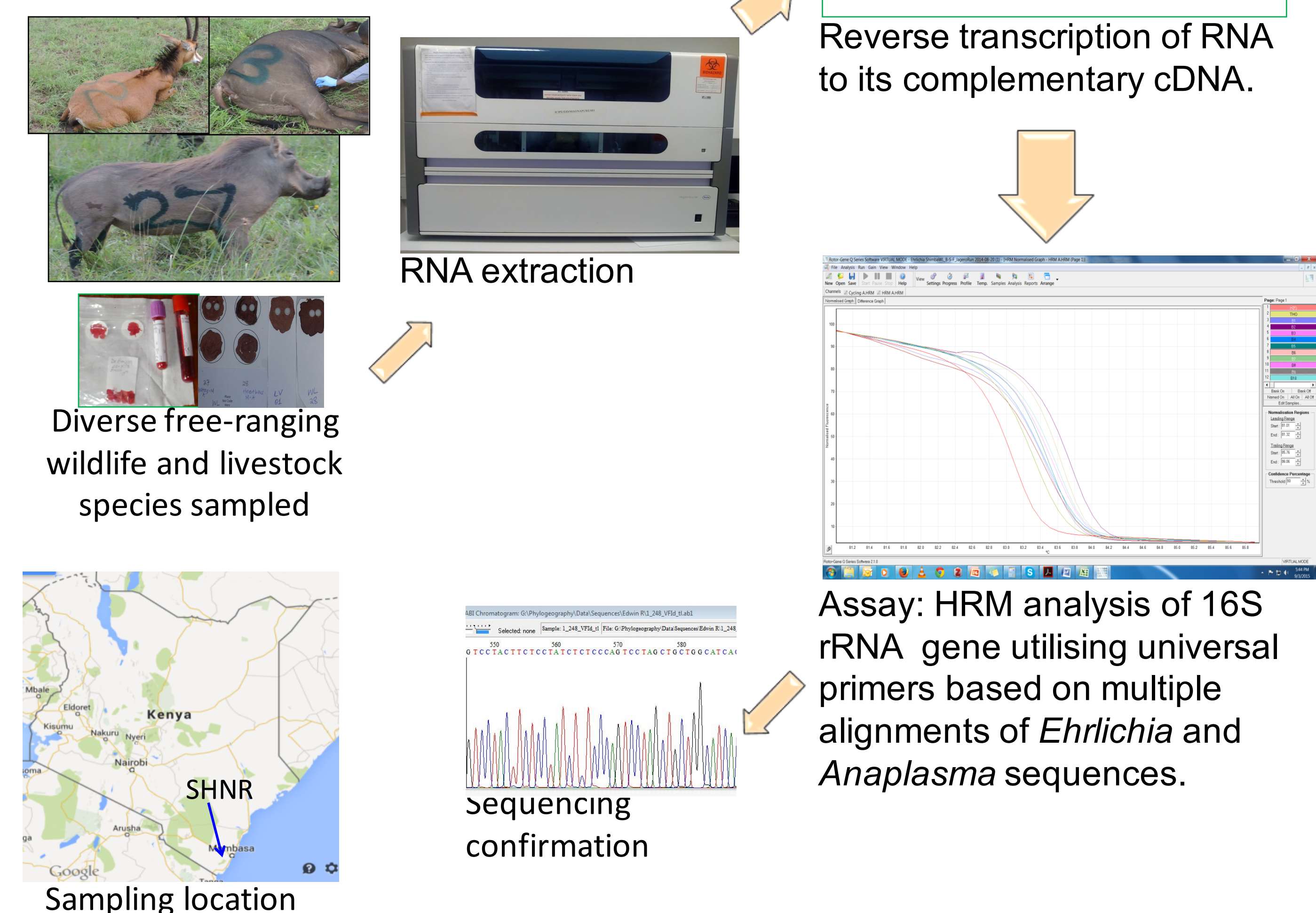
Geofrey Jagero^{1,3}, Daniel Ouso¹, Benedict Orindi¹, Edward Kariuki², Micky Mwamuye¹, Daniel Masiga¹, Louise Hamill³ and Jandouwe Villinger¹

¹International Centre of Insect Physiology and Ecology (*icipe*), Kenya; ²Kenya Wildlife Service, Kenya; ³The University of Edinburgh, UK
gjagero@icipe.org

INTRODUCTION

Anthropogenic activities remain key drivers of disease emergence as humans encroach into wildlife habitats that are major reservoirs of zoonotic bacterial pathogens. Little is known about the distribution and genetic characterisation of *Anaplasma* and *Ehrlichia*, the causative agents of human granulocytic anaplasmosis and human monocytic ehrlichiosis, respectively. We investigated these tick-borne zoonotic bacteria in wildlife and livestock in the Shimba Hills National Reserve (SHNR) of Kenya, an area in which wildlife live in close proximity to humans and livestock.

METHODS



CONCLUSION

The SHNR ecosystem provides an interface for wildlife-livestock interactions that may pose a challenge to food security in the region as well as conservation threats to the wildlife.

IMPACT

Diverse tick-borne bacteria pathogens were identified in wildlife and livestock populations, including emerging pathogens, such as *E. chaffeensis*, responsible for life-threatening human monocytic ehrlichiosis, as well as *Anaplasma marginale* and *A. phagocytophilum*, which can cause tick-borne fevers in both livestock and humans.

OBJECTIVES

To detect and identify tick-borne bacteria in wildlife and livestock in Shimba Hills National Reserve.

RESULTS

Table 1. Percent pathogens detected against the specimen storage method.

Pathogen detected	Wildlife					Livestock		
	Buffalo (n=18)	Sable antelope (n=5)	Warthog n=3	Bush-buck n=1	Hartebeest n=1	Cattle n=42	Sheep n=2	Goats n=17
<i>E. chaffeensis</i>	1 (5.6)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>E. spp</i>	2 (11.1)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>A. marginale</i>	4 (22.2)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (4.8)	0 (0.0)	3 (17.6)
<i>A. phagocytophilum</i>	0 (0.0)	1 (20.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Anaplasma spp.</i>	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (5.9)
Total	7 (38.9)	1 (20.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (4.8)	0 (0.0)	4 (23.5)

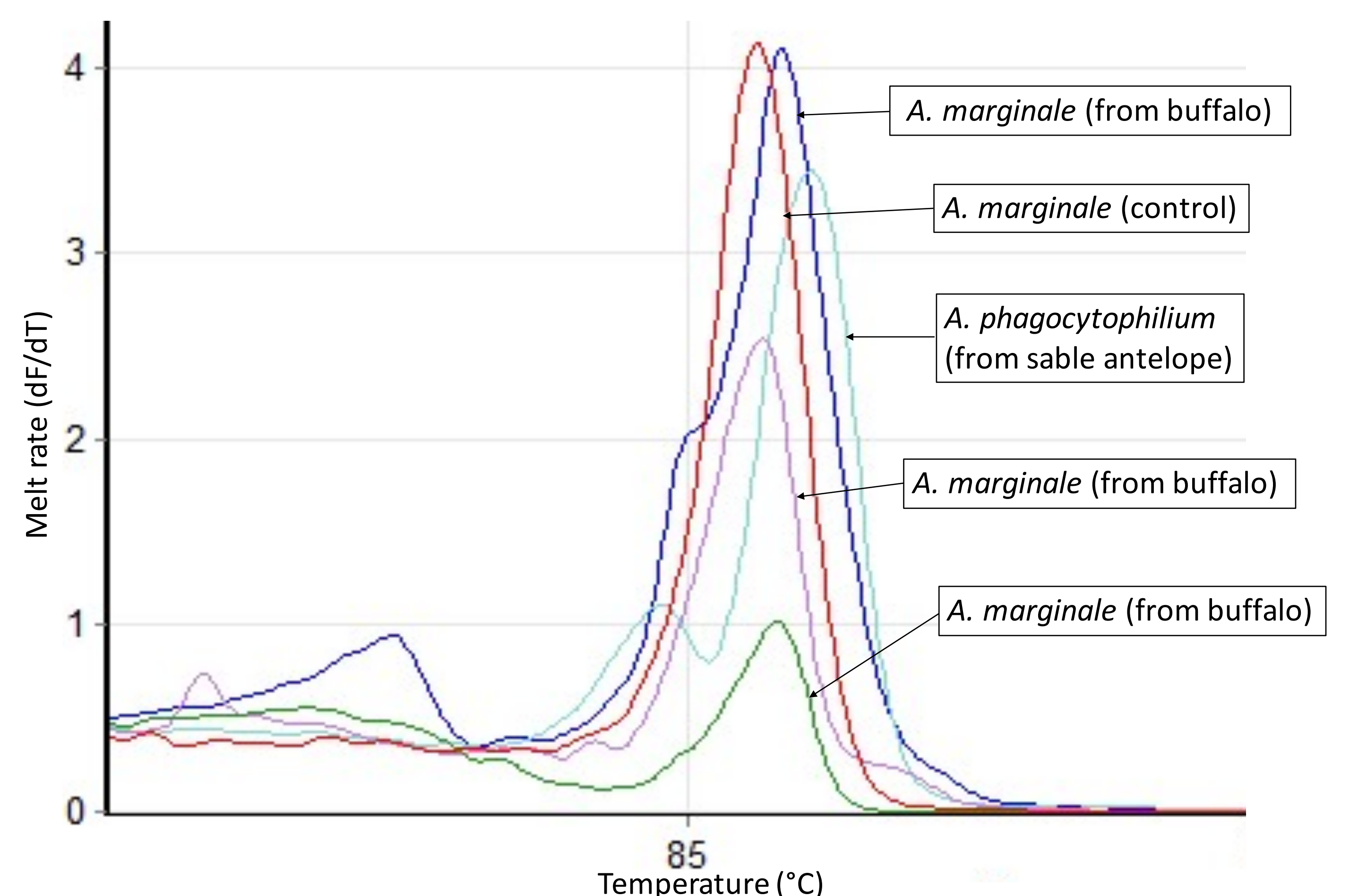


Figure 1: Melt rates (change in fluorescence with change in temperature) of *Anaplasma* spp. 16S PCR products from wildlife blood samples.

REFERENCES

- Chomel B.B., Belotto A. and Meslin F.-X. (2007) Wildlife, exotic pets, and emerging zoonoses. *Emerging Infectious Diseases* 13, 6–11.
- Smith K.F., Acevedo-Whitehouse K. and Pedersen A.B. (2009) The role of infectious diseases in biological conservation. *Animal Conservation* 12, 1–12.
- Taylor L.H., Latham S.M., Woolhouse M.E. (2001) Risk factors for human disease emergence. *Philosophical Transactions of the Royal Society London B: Biological Sciences* 356, 983–989.